

Molecular Genetics					
Identification number	Workload	Credit points	Term of studying	Frequency of occurrence	Duration
MN-B-SM (GB 1)	360 h	12 CP	1 <sup>st</sup> or 2 <sup>nd</sup> term of studying	Winter term, 2 <sup>nd</sup> half	7 weeks
<b>1</b>	<b>Type of lessons</b>		<b>Contact times</b>	<b>Self-study times</b>	<b>Intended group size*</b>
	a) Lectures		20 h	40 h	max. 8
	b) Practical/Lab		150 h	118 h	max. 2
	d) Seminar		8 h	24 h	max. 8
<b>2</b>	<b>Aims of the module and acquired skills</b> Students who successfully completed this module ... <ul style="list-style-type: none"> <li>• have acquired detailed knowledge of molecular genetics, the function of RNA-binding proteins and the different steps of eukaryotic gene expression, including pre-mRNA processing, RNA export, translation and RNA degradation.</li> <li>• have acquired experimental skills in state-of-the art methods in molecular biology and can and can independently design and perform small scientific projects related to the topics of the module.</li> <li>• have learned how to present research results in oral and written form and to critically discuss scientific publications related to the topic of the module on a professional level.</li> <li>• are able to transfer skills acquired in this module to other fields of biology.</li> </ul>				
<b>3</b>	<b>Contents of the module</b> <ul style="list-style-type: none"> <li>• Analysis of co- and post-transcriptional steps of human gene expression, with focus on regulation conferred by RNA-binding proteins</li> <li>• Applying recombinant DNA technologies, e.g. cloning, DNA preparation, etc.</li> <li>• Cell culture using immortalized human cell lines, transfection of plasmid DNA, expression of gene products (RNA/protein) and stable cell line generation</li> <li>• Functional characterization of RNA-binding proteins by siRNA-mediated knockdown and complementation assays</li> <li>• Extraction of nucleic acid and protein samples from cultured cells</li> <li>• Analysis of abundance and sub-cellular localization of proteins using immunofluorescence and western blotting</li> <li>• Techniques for monitoring alternative splicing and RNA degradation (RT-PCR, etc.)</li> <li>• Basic workflows for producing, analyzing and interpreting high-throughput RNA-sequencing data (focus on differential gene expression analysis, alternative splicing, isoform switches)</li> <li>• Addressing and solving scientific problems</li> </ul>				
<b>4</b>	<b>Teaching/Learning methods</b> <ul style="list-style-type: none"> <li>• Lectures; Practical/Lab (Project work);Seminar; Guidance to independent research; Training on presentation techniques in oral and written form</li> </ul>				
<b>5</b>	<b>Requirements for participation</b> Enrollment in the Master´s degree course "Biological Sciences" or in the Master´s degree course "Biochemistry"				

6	<p><b>Type of module examinations</b></p> <p>The final examination consists of three parts: Two hours written examination about topics of the lectures and the practical/lab part (50 % of the total module mark), oral presentation (25 % of the total module mark) and seminar paper (= short protocol in the form of a research paper about the project work; 25 % of the total module mark)</p>
7	<p><b>Requisites for the allocation of credits</b></p> <p>Regular and active participation Each examination part at least "sufficient" (see appendix of the examination regulations for details)</p>
8	<p><b>Compatibility with other Curricula</b></p> <p>Biological subject module in the Master´s degree course "Biochemistry"</p>
9	<p><b>Significance of the module mark for the overall grade</b></p> <p>In the Master´s degree course "Biological Sciences": 15 % of the overall grade (see also appendix of the examination regulations)</p>
10	<p><b>Module coordinator</b></p> <p>Prof. Dr. Niels Gehring, phone 470-3873, e-mail: ngehring@uni-koeln.de</p>
11	<p><b>Additional information</b></p> <p><b>Subject module</b> of the Master´s degree course "Biological Sciences", <b>Focus of research:</b> (G) Genetics and Cell Biology; (B) Biochemistry, Biotechnology and Biophysics <b>Participating faculty:</b> Prof. Dr. N. Gehring, Prof. Dr. D. Mörsdorf, Dr. G. Praefcke, Dr. V. Boehm <b>Literature:</b></p> <ul style="list-style-type: none"> <li>• Jocelyn E. Krebs, Elliott S Goldstein, Stephen T. Kilpatrick. Lewin's Genes XII, Jones and Bartlett Publishers, Inc; 12th Revised edition edition (2017), Part III and IV</li> <li>• James D. Watson, Tania A. Baker, Alexander Gann, Michael Levine, Richard Losick. Molecular Biology of the Gene, Seventh Edition (2013), Part 4 and 5</li> <li>• Additional subject-specific literature will be provided at the beginning of the module</li> </ul> <p><b>General time schedule:</b> Week 1-6 (Mon.-Fri.): Lectures, Practical/Lab (Project work); (daily from approximately 9 a.m. to 5 p.m. including lunch break, times may vary depending on project's tasks) as well as preparation for the seminar talk (held at the end of week 6); Week 7 (Mon.-Thu.): Preparation for the written examination</p> <p><b>Note:</b> The module contains hand-on laboratory work conducted individually or by small groups of students and is taught mainly in course rooms. The module does contain computer-based practicals/research as a minor component. The schedule and content of the module have changed compared to previous years.</p> <p><b>Introduction to the module:</b> December 2, 2019 at 9 a.m., Center for Molecular Biosciences (COMB), seminar room 0.46 (ground floor)</p> <p><b>Written examination:</b> January 31, 2020, second/supplementary examination March 20, 2020; the latter date may vary if students and module coordinator agree. More details will be given at the beginning of the module.</p>

\*7 students from the Master´s degree course "Biological Sciences" and 1 student from the Master´s degree course "Biochemistry".